Chiral Bicyclic Guanidine-Catalyzed Enantioselective Sulfenylation of Oxindoles and Benzofuran-2(3*H*)-ones

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Supporting Information



ABSTRACT: A chiral bicyclic guanidine-catalyzed enantioselective sulfenylation of 3-substituted oxindoles to N-(sulfanyl)-succinimides has been developed. A series of unprecedented 3-sulfenylated oxindoles, such as 3-benzyl/alkyl-substituted 3-benzyl/alkyloxindoles, were obtained with high enantioselectivities (up to 98% ee). This methodology is also effective for the first asymmetric sulfenylation of benzofuran-2(3H)-ones, providing 3-benzyl-3-benzylthio-substituted benzofuran-2(3H)-ones with satisfactory results (up to 95% ee).

3,3-Disubstituted oxindoles, featuring a quaternary or heteroquaternary stereogenic center on the C3 position, are one of the most common structural scaffolds in bioactive natural and non-natural products.¹ Hence, asymmetric synthesis of 3,3disubstituted oxindoles has attracted significant attention from chemists during the past few decades.² In particular, the direct asymmetric reaction of prochiral 3-substituted oxindoles to electrophiles has been demonstrated as a powerful strategy to access these important chiral molecules.² For example, asymmetric sulfenylation of 3-substituted oxindoles has recently been devised to furnish directly stereoselective implantation of sulfur onto the C3 position of oxindoles.³⁻⁵ In 2012, Feng and co-workers presented pioneering work on asymmetric sulfenylation of 3-substituted oxindoles through a cooperative catalysis of achiral a Brønsted base and chiral N₁N'-dioxide-Sc(OTf)₃ complex, leading to 3-arylthio-substituted 3-aryl/ benzyl/alkyloxindoles with excellent enantioselectivities (Scheme 1a).^{5a} Almost simultaneously, the Enders group reported an organocatalytic example with the same substrate scope as the Feng's.^{5b} Li, Cheng and co-workers described a highly enantioselective sulfenylation to build 3-arylthiosubstituted 3-aryl/alkyloxindoles (Scheme 1b).^{5c} Our group developed an asymmetric organocatalytic variant of 3aryloxindoles to access chiral 3-arylthio-, 3-benzylthio-, and 3alkylthio-substituted 3-aryloxindoles in satisfactory results (Scheme 1c).^{5d} Nevertheless, to date to the best of our

knowledge, four kinds of 3-sulfenylated oxindoles, i.e. 3-benzyl/ alkyl-substituted 3-benzyl/alkyloxindoles, still remain unexploited (Scheme 1d). In our preliminary work, no reaction was observed between 3-benzyl-substituted oxindole and *N*-(benzylthio)succinimide using our previously reported reaction conditions^{5d} (Scheme 1e). Thus, enantioselectivity and the poor reactivity represent the important yet difficult challenges for these unmet asymmetric sulfenylation of 3-substituted oxindoles.

In recent years, we were keen on the development of asymmetric organocatalytic methodologies to synthesize various significant molecules which contain quaternary and heteroquaternay stereogenic centers.^{4c,d,5d,6} Among these works, chiral C_2 -symmetric bicyclic guanidines exhibited satisfactory ability in stereoselective control toward several reactions using 3-benzyl-substituted oxindoles as nucleophiles, such as 1,4-conjugate addition to *N*-maleimides^{6a} and 2-cyclopentenone^{6e} as well as alkylation.^{6f} Owing to the similarity in structural frameworks between *N*-succinimides and *N*-maleimides, it is plausible that bicyclic guanidine catalysts could lead to similar enantioselective outcomes for these two kinds of electrophiles. More importantly, chiral bicyclic

Received: July 12, 2015 Published: August 6, 2015 Scheme 1. Established Asymmetric Sulfenylation of 3-Substituted Oxindoles and the Remaining Challenges

Previous works (accomplished scopes) a) Feng's work:^{5a} L*-Sc(OTf)₃, Na₂CO₃, CH₂Cl₂, 35 °C (L* = chiral *N*,*N*'-dioxide ligand) Enders' work:^{5b} chiral squaramide-tertiary amine, CHCl₃, rt



b) Li and Cheng's work:^{4c} quinidine, CH₂Cl₂, -80 °C



c) Our previous work:^{4d} (DHQD)₂PHAL, *m*-xylene, 30 °C



d) Unexploited sulfenylated products



guanidines as superbases have been demonstrated as competent catalysts in several sluggish reactions.^{7,8} In this regard, we envisioned that chiral bicyclic guanidines should be feasible to accomplish the desired sulfenylation.

To prove the feasibility of the guanidine catalyst, we first attempted an achiral approach of sulfenylation between 3benzyl-substituted oxindole 1a and N-(benzylthio)succinimide 2a by employing 10 mol % of 1,1,3,3-tetramethyl guanidine (TMG) as a catalyst in toluene at 30 °C. The reaction completed within 12 h to afford the sulfenylated product 4aa in 83% yield (Table 1, entry 1). The positive results allowed us to attempt chiral bicyclic guanidines. To our delight, we found that 10 mol % of bicyclic guanidine I, which was derived from L-tert-leucine, could promote the reaction; the desired adduct 4aa was obtained in 53% yield with 61% ee after 28 h (entry 2). Afterward, the solvent effect was tested (entries 3-9). It was found that cyclopentyl methyl ether (CPME) was the most suitable solvent for providing the best enantioselectivity (73% ee, entry 8). The lower temperature led to an increase of the enantioselectivity level (entries 10-13), and -10 °C was proven to be the best for producing 4aa in 56% yield with 82% ee (entry 12).

Since the side chain would probably affect the enantioselectivity, we were engaged to prepare and test a series of other chiral bicyclic guanidines II–V with distinct side chains (entries 14–17). Bicyclic guanidine II with isopropyl as the side chain was discovered to significantly improve the ee value of 4aa (50% yield, 90% ee, entry 14). The effect of *N*-substituents of 3-oxidoles was then evaluated using 10 mol % of bicyclic guanidine II at –10 °C in CPME (entries 18–20). The results described that the oxindole 1b with methyl as the *N*-substituent group gave the corresponding adduct **4ba** in 45% yield with 92% ee (entry 18). The reactivity was further improved by increasing the amount of catalyst **II** from 0.1 to 0.2 equiv, and **4ba** was obtained in 82% yield with 90% ee after 37 h (entry 21).

With the optimal conditions in hand, the substrate scope, involving various 3-benzyl and alkyl-substituted oxindoles 1 and N-(benzyl/alkylthio)succinimides 2, was investigated (Table 2 and Figure 1). First, we performed the reactions of N-(benzylthio)succinimide 2a with various 3-benzyl-substituted oxindoles (1e-1n) (entries 1-10). The results showed that the reactions proceeded smoothly and gave the expected adducts (4ea-4na) in 75-99% yields with 90-98% ee within 37-96 h (entries 1-10). Although the position and electronic properties of the substituents on the aromatic ring of benzyl groups at the C3-position of oxindoles 3 did not affect the ee value, methoxyl as the electron-donating group was found to decrease the reactivity (entries 5-6). Moreover, the substituent groups on the C4, C5, C6, and C7 positions of the aromatic ring of oxindoles 1 made the reaction sluggish (entries 7-10).

Meanwhile, the sulfenylation reactions of 3-benzyl-substituted oxindole 1b with different *N*-(benzylthio)succinimides 2b-g were conducted, providing the corresponding adducts 4bb-bg in 76–86% yield with 90–95% ee within 63–96 h (entries 11–16). The reaction between oxindole 1b and *N*-(cyclohexylthio)succinimide 2h was found to produce 3alkylthio-substituted 3-benzyloxindole 4bh in moderate yield and good enantioselectivity (entry 17). Satisfactory results could also be attained for the reactions between 3-alkylsubstituted oxindoles 1o-p and *N*-(benzylthio)succinimides (2a-b and 2e) (entries 18-20). Finally, the achieved 3cyclohexylthio-substituted 3-methyloxindole 4oh with 65% yield and 92% ee indicated the feasibility of this protocol for the highly enantioselective reaction of 3-alkyl-substituted oxindoles with *N*-(alkylthio)succinimides (entry 21).

In order to illustrate the versatility of this protocol, we were intrigued to examine the sulfenylation of 3-substituted benzofuran-2(3H)-ones, mainly due to their analogous structural features to 3-substituted oxindoles as well as the significant potentials of 3,3-disubstituted benzofuran-2(3H)ones in organic and medicinal chemistry.9-14 Indeed, several asymmetric reactions of 3-substituted benzofuran-2(3H)-ones have been disclosed to date, such as Steglich rearrangement,¹⁰ 1,4-conjugate addition,¹¹ fluorination,¹² amination,¹³ and allylic alkylation;¹⁴ however, no example of asymmetric sulfenylation has yet been reported. The reactions of 3-substituted benzofuran-2(3H)-ones 3 with N-(benzylthio)succinimide 2a or N-(*n*-propylthio)succinimide 2i were investigated in the presence of 10 mol % of bicyclic guanidine I in MTBE as solvent at -10 °C (Scheme 2).¹⁵ The results indicated that 3substituted benzofuran-2(3H)-ones 3 have higher reactivity than 3-substituted oxindoles 2 for the remarkably shortened reaction time (3-5 h). The reaction conditions were suitable to introduce a benzylthio group onto benzofuran-2(3H)-ones; the corresponding adducts 5aa-5ea and 5fa were obtained in 68-93% yield with 71-95% ee. Moderate enantioselectivities (76 and 74% ee) were observed in the creation of 3-n-propylthiosubstituted benzofuran-2(3H)-ones (**5ai** and **5fi**). The absolute configurations of sulfenylated products 4 and 5 were assigned based on X-ray crystallographic analysis of a single crystal of 5aa.¹⁶

Next, the synthetic transformation of sulfenylated adducts was attempted to demonstrate the utility of the established

Table 1. Optimization of the Reaction Conditions⁴



entry	1	catalyst	solvent	T (°C)	<i>t</i> (h)	4	yield ^b (%)	ee ^c (%)
1	1a	TMG	toluene	30	12	4aa	83	N.D. ^d
2	1a	Ι	toluene	30	28	4aa	53	61
3	1a	Ι	CH_2Cl_2	30	14	4aa	32	35
4	1a	Ι	THF	30	28	4aa	N.P. ^e	N.D. ^d
5	1a	Ι	Et ₂ O	30	14	4aa	59	63
6	1a	Ι	CH ₃ CN	30	3	4aa	40	20
7	1a	I	hexane	30	14	4aa	32	50
8	1a	Ι	CPME	30	12	4aa	42	73
9	1a	Ι	TBME	30	12	4aa	28	67
10	1a	Ι	CPME	10	12	4aa	34	81
11	1a	Ι	CPME	0	18	4aa	58	81
12	1a	Ι	CPME	-10	24	4aa	56	82
13	1a	I	CPME	-20	24	4aa	52	81
14	1a	II	CPME	-10	24	4aa	50	90
15	1a	III	CPME	-10	24	4aa	49	87
16	1a	IV	CPME	-10	24	4aa	40	87
17	1a	V	CPME	-10	24	4aa	N.R. ^{<i>f</i>}	N.D. ^d
18 ^g	1b	II	CPME	-10	24	4ba	45	92
19	1c	II	CPME	-10	24	4ca	39	90
20	1d	II	CPME	-10	24	4da	15	79
21 ^{<i>h</i>}	1b	II	CPME	-10	37	4ba	82	90

^{*a*}The reaction was carried out with 0.05 mmol of 1, 0.06 mmol of 2a, and 0.05 mmol of catalyst in 0.5 mL of solvent. ^{*b*}Isolated yield. ^{*c*}Determined by HPLC methods. ^{*d*}N.D. = not determined. ^{*e*}N.P. = no desired product. ^{*f*}N.R. = no reaction. ^{*g*}0.1 mmol scale, 94 h, 84% yield, 92% ee. ^{*h*}20 mol % of catalyst was used, 0.1 mmol scale.

methodology (Figure 2). It was found that the adduct 3be could be conveniently oxidized by *m*CPBA in dicholoromethane to provide the sulfone 4 in 95% yield and without compromising the ee value. Subsequently, based on the observed stereochemistry of sulfenylated adducts, a reasonable side-on transition state (TS) was proposed in Figure 2.^{6a} The benzyl or alkyl groups of nucleophiles 1 and 3 have longer or bulkier carbon chains than aryl groups, which leads to stronger shielding effects from the side chain of chiral guanidines (*tert*-butyl for catalyst I and isopropyl for catalyst II), thus presenting higher enantioselective results.

In summary, we have developed two highly enantioselective sulfenylation reactions of oxindoles and benzofuran-2(3H)-ones to *N*-(sulfanyl)succinimides, respectively. In the presence of a chiral bicyclic guanidine as the catalyst, the reaction between 3-benzyl/alkyl-substituted oxindoles and *N*-(benzyl/ alkylthio)succinimides could work smoothly, affording a series of unprecedented sulfenylated adducts, including 3-benzyl/ alkyl-substituted 3-benzyl/alkyloxindoles, in good to excellent yields and enantioselectivities (up to 98% ee). This work successfully replenishes the desired substrate scope in asymmetric sulfenylation of 3-substituted oxindoles. Furthermore, the first asymmetric sulfenylation of benzofuran-2(3H)-ones has been accomplished upon this developed methodology,

to afford biologically important 3-benzyl-3-benzylthio-substituted benzofuran-2(3H)-ones with satisfactory results (up to 95% ee).

EXPERIMENTAL SECTION

General Information. General Procedures and Methods. Experiments involving moisture and/or air sensitive components were performed under a positive pressure of nitrogen in oven-dried glassware equipped with a rubber septum inlet. Dried solvents and liquid reagents were transferred by oven-dried syringes or hypodermic syringe cooled to ambient temperature in a desiccator. Reactions mixtures were stirred in a 4 mL sample vial with Teflon-coated magnetic stirring bars unless otherwise stated. Moisture in nonvolatile reagents/compounds was removed in high *vacuo* by means of an oil pump and subsequent purging with nitrogen. Solvents were removed in vacuo under ~30 mmHg and heated with a water bath at 30-35 °C using a rotary evaporator with an aspirator.

All experiments were monitored by analytical thin layer chromatography (TLC). TLC was performed on precoated plates, 60 F_{254} . After elution, the plate was visualized under UV illumination at 254 nm for UV active material. Further visualization was achieved by staining KMnO₄, ceric molybdate, or anisaldehyde solution. For those using the aqueous stains, the TLC plates were heated on a hot plate.

Columns for flash chromatography (FC) contained silica gel 200– 300 mesh. Columns were packed as a slurry of silica gel in petroleum Table 2. Variation of Oxindoles 1 and N-(Sulfanyl)succinimides 2^a

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	R1		ŝ	`N ^{≤⊥}	Ň H II	R ¹ SR ³
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₩- <u></u> ,	~n		\mathcal{Y}	CPME,	,-10 °C स⁻म्	∕_n∕─∪
	Ме 1		0 2			Me 4
entry	1	2	<i>t</i> (h)	4	yield (%) ^b	ee (%) ^c
1	1e	2a	50	4ea	99	92
2	1f	2a	37	4fa	89	93
3 ^d	1g	2a	37	4ga	99	93
4 ^e	1h	2a	37	4ha	95	92
5	1i	2a	72	4ia	75	95
6	1j	2a	96	4ja	82	90
7	1k	2a	96	4ka	83	92
8	11	2a	96	4la	75	90
9	1m	2a	72	4ma	97	98
10	1n	2a	96	4na	85	90
11^{f}	1b	2b	72	4bb	77	90
12	1b	2c	96	4bc	84	90
13	1b	2d	96	4bd	86	95
14 ^g	1b	2e	63	4be	83	90
15	1b	2f	63	4bf	78	92
16	1b	2g	72	4bg	76	92
17	1b	2h	96	4bh	65	86
18	10	2a	71	40a	85	90
19	1p	2a	96	4pa	70	92
20	10	2b	71	4ob	81	92
21	10	2e	71	40e	93	92
22	10	2h	96	4oh	65	92

^{*a*}The reaction was carried out with 0.1 mmol of 1, 0.12 mmol of 2 and 0.02 mmol of catalyst in 1.0 mL solvent. ^{*b*}Isolated yield. ^{*c*}Determined by HPLC methods. ^{*d*}10 mol % of II, 72 h, 99% yield, 93% ee. ^{*e*}10 mol % of II, 84 h, 80% yield, 92% ee. ^{*f*}10 mol % of II, 72 h, 70% yield, 90% ee. ^{*g*}10 mol % of II, 72 h, 75% yield, 90% ee.



Figure 1. Structures of substrates.

ether and equilibrated solution using the appropriate solvent system. The elution was assisted by applying a pressure of about 2 atm with an air pump.

Instrumentation. Proton nuclear magnetic resonance (¹H NMR) and carbon NMR (¹³C NMR) spectra were recorded in CDCl₃ unless otherwise stated. ¹H (300 MHz) and ¹³C (75 MHz) NMR were performed on a 300 MHz spectrometer. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl₃ (¹H NMR: δ 7.26, singlet; ¹³C NMR: δ 77.0, triplet). Multiplicities were given as *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in hertz (Hz). The number of proton atoms (*n*) for a given

Note



^{*a*}The reaction was carried out with 0.1 mmol of 3, 0.15 mmol of 2, and 0.01 mmol of catalyst in 3.0 mL of MTBE at -10 °C (MTBE = methyl *tert*-butyl methyl ether). Yield of isolated product. Ee's were determined by HPLC analysis on a chiral stationary phase. ^{*b*}1.0 mL of MTBE was used.



Figure 2. Synthetic transformation of sulfenylated product 4ae to sulfone 6 and the plausible transition-state model.

resonance was indicated by *n*H. The number of carbon atoms (*n*) for a given resonance was indicated by *n*C. HRMS (Analyzer: TOF) was reported in units of mass of charge ratio (*m/z*). Mass samples were dissolved in CH₃CN (HPLC grade) unless otherwise stated. Optical rotations were recorded on a polarimeter with a sodium lamp (wavelength = 589 nm) and reported as follows: $[\alpha]_{\lambda}^{T \circ C}$ (*c* = g/100 mL, solvent). Melting points were determined on a melting point apparatus.

Enantiomeric excesses were determined by chiral High Performance Liquid Chromatography (HPLC) analysis. UV detection was monitored at 254, 230, and 210 nm at the same time. HPLC samples

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were dissolved in HPLC grade isopropanol (IPA) unless otherwise stated.

Materials. All commercial reagents were purchased with the highest purity grade. They were used without further purification unless specified. All solvents used, mainly petroleum ether (PE) and ethyl acetate (EtOAc), were distilled. Anhydrous CH_2Cl_2 was freshly distilled from CaH_2 and stored under a N_2 atmosphere. Toluene, hexane, diethyl ether, CH_3CN , CPME, TBME, and THF were freshly distilled from sodium/benzophenone before use. All compounds synthesized were stored in a 4 °C freezer, and light-sensitive compounds were protected with aluminum foil.

General Procedure 1. *N*-Methyl-3-benzyl-2-oxindoles **1** (0.1 mmol, 1.0 equiv) and guanidine **II** (0.02 mmol, 0.2 equiv) were dissolved in cyclopentyl methyl ether (1.0 mL) and stirred at -10 °C for 5 min. Then *N*-(sulfanyl)succinimides **2** (0.12 mmol, 1.2 equiv) were added. The reaction mixture was stirred at -10 °C and monitored by TLC. Upon complete consumption of **1**, the reaction mixture was directly loaded onto a short silica gel column, followed by gradient elution with petroleum ether/ethyl acetate (10/1-5/1 ratio). Removing the solvent *in vacuo* afforded products **3**.

General Procedure 2. Benzofuran-2(3*H*)-ones 3 (0.1 mmol, 1.0 equiv) and guanidine I (0.01 mmol, 0.1 equiv) were dissolved in methyl *tert*-butyl ether (3.0 mL) and stirred at -10 °C for 5 min. Then *N*-(sulfanyl)succinimides 2 (0.15 mmol, 1.5 equiv) were added. The reaction mixture was stirred at -10 °C and monitored by TLC. Upon complete consumption of 3, the reaction mixture was directly loaded onto a short silica gel column, followed by gradient elution with petroleum ether/ethyl acetate (10/1-4/1 ratio). Removing the solvent *in vacuo* afforded products 5.

4ba, (5)-(–)-3-Benzyl-3-(benzylthio)-1-methylindolin-2-one. Colorless oil, 30 mg (0.1 mmol), 82% yield, 90% ee. $[\alpha]_D^{26}$ –37.0 (*c* 0.63, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.32 (dd, *J* = 7.4, 0.7 Hz, 1H), 7.24–7.00 (m, 10H), 6.88 (dd, *J* = 7.3, 2.1 Hz, 2H), 6.56 (d, *J* = 7.7 Hz, 1H), 3.70–3.58 (m, 2H), 3.41 (d, *J* = 13.1 Hz, 1H), 3.27 (d, *J* = 13.1 Hz, 1H), 2.84 (s, 3H). ¹³CNMR (75 MHz, CDCl₃) δ 176.0, 143.3, 136.7, 134.7, 130.1, 129.0 (two peaks), 128.3, 127.7, 127.2, 126.9, 124.7, 122.5, 108.0, 56.3, 42.2, 34.0, 26.0 HRMS (ESI) *m/z* 360.1421 (M+H⁺), calcd for C₂₃H₂₁NOS 360.1422. The *ee* was determined by HPLC analysis. CHIRALPAK AD-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 210 nm; retention time: 14.2 min (major) and 11.1 min (minor).

4ea, (5)-(-)-3-(Benzylthio)-3-(4-fluorobenzyl)-1-methylindolin-2-one. Colorless oil, 37 mg (0.1 mmol), 99% yield, 92% ee. [α]₂₆²⁶ -29.5 (*c* 0.62, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.33 (d, *J* = 7.4 Hz, 1H), 7.25-7.03 (m, 7H), 6.88-6.79 (m, 2H), 6.72 (t, *J* = 8.4 Hz, 2H), 6.58 (d, *J* = 7.8 Hz, 1H), 3.69-3.58 (m, 2H), 3.38 (d, *J* = 13.1 Hz, 1H), 3.23 (d, *J* = 13.1 Hz, 1H), 2.83 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 175.9, 163.49, 160.2, 143.3, 136.6, 131.6 (two peaks), 130.4 (two peaks), 129.0 (two peaks), 128.2 (two peaks), 127.1, 124.5, 122.6, 114.7, 114.4, 108.2, 56.2, 41.4, 34.0, 26.0. HRMS (ESI) *m/z* 378.1322 (M+H⁺), calcd for C₂₃H₂₀FNOS 378.1328. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.7 min (major) and 5.4 min (minor).

4fa, (S)-(–)-3-(Benzylthio)-3-(3-fluorobenzyl)-1-methylindolin-2-one. Colorless oil, 34 mg (0.1 mmol), 89% yield, 93% ee. [α]₂₆²⁶ –67.3 (*c* 0.39, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.32 (dd, *J* = 7.4, 0.6 Hz, 1H), 7.25–6.97 (m, 8H), 6.76 (td, *J* = 8.4, 2.0 Hz, 1H), 6.69 (d, *J* = 7.7 Hz, 1H), 6.62–6.55 (m, 2H), 3.70–3.61 (m, 2H), 3.41 (d, *J* = 13.1 Hz, 1H), 3.26 (d, *J* = 13.1 Hz, 1H), 2.86 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 175.8, 163.4, 160.9, 143.3, 137.2 (two peaks), 136.6, 129.1 (two peaks), 128.3, 127.9, 127.2, 125.9 (two peaks), 124.5, 122.7, 117.0, 116.8, 113.9, 113.7, 108.2, 56.0, 41.8, 34.0, 26.1. HRMS (ESI) *m/z* 378.1324 (M+H⁺), calcd for C₂₃H₂₀FNOS 378.1328. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.38 min (major) and 5.74 min (minor).

4ga, (S)-(-)-3-(Benzylthio)-1-methyl-3-(3-(trifluoromethyl)benzyl)indolin-2-one. Colorless oil, 42 mg (0.1 mmol), 99% yield, 93% ee. $[\alpha]_{D}^{26}$ –45.8 (*c* 0.71, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.32 (d, *J* = 7.4 Hz, 2H), 7.26–7.03 (m, 10H), 6.55 (d, *J* = 7.8 Hz, 1H), 3.66 (s, 2H), 3.45 (d, *J* = 13.1 Hz, 1H), 3.29 (d, *J* = 13.1 Hz, 1H), 2.80 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 175.7, 143.1, 136.5, 135.6, 133.6, 129.1 (two peaks), 128.2 (two peaks), 127.6, 127.2, 126.7 (two peaks), 124.5, 123.6, 122.7, 108.2, 56.1, 42.1, 34.1, 25.9. HRMS (ESI) *m*/*z* 428.1295 (M+H⁺), calcd for C₂₄H₂₀F₃NOS 428.1296. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 4.9 min (major) and 4.6 min (minor).

4ha, **(S)-(–)-3-(Benzylthio)-3-(2-fluorobenzyl)-1-methylindolin-2-one.** Colorless oil, 36 mg (0.1 mmol), 95% yield, 92% ee. $[α]_{26}^{D}$ –35.4 (*c* 0.64, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.30 (d, *J* = 7.4 Hz, 1H), 7.24–7.00 (m, 9H), 6.93–6.84 (m, 1H), 6.77 (t, *J* = 9.0 Hz, 1H), 6.59 (d, *J* = 7.8 Hz, 1H), 3.73–3.63 (m, 2H), 3.53 (d, *J* = 13.4 Hz, 1H), 3.36 (d, *J* = 13.4 Hz, 1H), 2.94 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 162.4, 159.2, 143.02, 136.6, 131.7 (two peaks), 128.9 (two peaks), 128.3, 127.8, 127.1, 125.0 (two peaks), 123.5 (two peaks), 122.6, 122.4 (two peaks), 115.2, 114.9, 107.8, 55.7, 33.9, 26.2. HRMS (ESI) *m/z* 378.1332 (M+H⁺), calcd for C₂₃H₂₀FNOS 378.1328. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.7 min (major) and 5.4 min (minor).

4ja, **(S)-(-)-3-(Benzylthio)-3-(2-methoxybenzyl)-1-methylindolin-2-one.** Colorless oil, 32 mg (0.1 mmol), 82% yield, 90% ee. $[\alpha]_{2^6}^{2^6}$ -67.7 (*c* 0.54, CHCl₃). ¹H NMR (400 MHz, CDCl₃) δ 7.25-7.11 (m, 7H), 7.08-7.01 (m, 2H), 6.96 (td, *J* = 7.6, 0.8 Hz, 1H), 6.71 (td, *J* = 7.5, 0.9 Hz, 1H), 6.58-6.55 (m, 2H), 3.67 (d, *J* = 12.5 Hz, 1H), 3.62 (d, *J* = 12.9 Hz, 2H), 3.53 (s, 3H), 3.33 (d, *J* = 13.3 Hz, 1H), 2.96 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 176.7, 157.3, 143.1, 136.8, 131.2, 129.1, 128.3 (two peaks), 127.0, 125.5, 123.9, 121.8, 119.8, 109.9, 107.5, 56.2, 54.7, 34.7, 34.0, 26.2. HRMS (ESI) *m/z* 390.1521 (M+H⁺), calcd for C₂₄H₂₃NO₂S 390.1528. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 nm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 24.0 min (major) and 21.1 min (minor).

4ka, **(5)-(-)-3-Benzyl-3-(benzylthio)-1,4-dimethylindolin-2**one. Colorless oil, 32 mg (0.1 mmol), 83% yield, 92% ee. $[\alpha]_D^{26}$ -14.5 (*c* 0.88, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.24–6.96 (m, 10H), 6.88 (dd, *J* = 7.0, 2.2 Hz, 2H), 6.46 (d, *J* = 7.9 Hz, 1H), 3.71– 3.59 (m, 2H), 3.39 (d, *J* = 13.1 Hz, 1H), 3.25 (d, *J* = 13.1 Hz, 1H), 2.83 (s, 3H), 2.35 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 175.9, 140.9, 136.8, 134.8, 132.1, 130.2, 129.1 (two peaks), 128.3, 127.7, 127.1, 126.8, 125.3, 107.7, 56.4, 42.2, 34.0, 26.1, 21.2. HRMS (ESI) *m*/ *z* 373.1505 (M+H⁺), calcd for C₂₄H₂₃NOS 373.1500. The *ee* was determined by HPLC analysis. LUX AMYLOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 16.4 min (major) and 14.8 min (minor).

4la, (5)-(–)-3-Benzyl-3-(benzylthio)-1,5-dimethylindolin-2one. Colorless oil, 28 mg (0.1 mmol), 75% yield, 90% ee. $[\alpha]_D^{26}$ – 49.95 (*c* 0.47, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.25–6.96 (m, 10H), 6.89 (d, *J* = 5.4 Hz, 2H), 6.46 (d, *J* = 7.6 Hz, 1H), 3.72–3.58 (m, 2H), 3.39 (d, *J* = 12.9 Hz, 1H), 3.25 (d, *J* = 13.5 Hz, 1H), 2.83 (s, 3H), 2.35 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 175.9, 141.0, 136.8, 134.8, 132.1, 130.2, 129.1 (two peaks), 128.3, 127.7, 127.1, 126.8, 125.3, 107.7, 56.4, 42.2, 34.0, 26.1, 21.2. HRMS (ESI) *m/z* 373.1505 (M+H⁺), calcd for $C_{24}H_{23}NOS$ 373.1500. The *ee* was determined by HPLC analysis. CHIRALCEL OD-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.7 min (major) and 18.4 min (minor).

4ma, (S)-(-)-3-Benzyl-3-(benzylthio)-6-fluoro-1-methylindolin-2-one. Colorless oil, 36 mg (0.1 mmol), 97% yield, 98% ee. $[\alpha]_{26}^{26}$ -92.2 (*c* 0.24, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.29-6.99 (m, 9H), 6.87 (d, *J* = 5.9 Hz, 2H), 6.75 (t, *J* = 8.8 Hz, 1H), 6.29 (d, *J* = 8.8 Hz, 1H), 3.67 (s, 2H), 3.40 (d, *J* = 13.1 Hz, 1H), 3.24 (d, *J* = 13.1 Hz, 1H), 2.80 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.3, 165.0, 161.7, 144.7 (two peaks), 136.5, 134.5, 130.1, 129.0, 128.3, 127.8, 127.1 (two peaks), 125.8 (two peaks), 123.6 (two peaks), 108.8, 108.5, 97.0, 96.6, 55.8, 42.3, 34.1, 26.2. HRMS (ESI) *m/z* 377.1252 (M+H⁺), calcd for C₂₃H₂₀FNOS 377.1250. The *ee* was determined by HPLC analysis. CHIRALPAK ID-3 (3.0 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.0 min (major) and 11.3 min (minor).

4na, (5)-(–)-3-Benzyl-3-(benzylthio)-7-fluoro-1-methylindolin-2-one. Colorless oil, 32 mg (0.1 mmol), 85% yield, 90% ee. $[\alpha]_{26}^{D}$ –49.98 (*c* 0.14, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.20 (dt, *J* = 6.7, 5.8 Hz, 3H), 7.10 (ddd, *J* = 15.7, 8.0, 4.2 Hz, 6H), 7.04– 6.90 (m, 2H), 6.87 (dd, *J* = 7.5, 1.8 Hz, 2H), 3.67 (s, 2H), 3.42 (d, *J* = 13.1 Hz, 1H), 3.24 (d, *J* = 13.1 Hz, 1H), 3.03 (d, *J* = 2.6 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 175.7, 136.4, 134.3, 131.3, 130.1, 129.0, 128.3, 127.8, 127.1 (two peaks), 123.0 (two peaks), 120.5, 116.7 (two peaks), 56.4, 42.5, 34.2, 28.5 (two peaks). HRMS (ESI) *m/z* 377.1252 (M+H⁺), calcd for C₂₃H₂₀FNOS 377.1250. The ee was determined by HPLC analysis. CHIRALPAK ID-3 (4.6 mm i.d. × 250 mm); hexane/ 2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 12.7 min (major) and 14.7 min (minor).

4bb, **(5)**-(–)-**3**-**Benzyl-3**-((**4**-fluorobenzyl)thio)-1-methylindolin-2-one. Colorless oil, 29 mg (0.1 mmol), 77% yield, 90% ee. $[\alpha]_{D}^{26}$ –43.8 (*c* 0.43, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.31 (d, *J* = 7.4 Hz, 1H), 7.21 (t, *J* = 7.7 Hz, 1H), 7.07 (dd, *J* = 13.7, 6.2 Hz, 6H), 6.87 (dd, *J* = 10.4, 4.9 Hz, 4H), 6.56 (d, *J* = 7.8 Hz, 1H), 3.68– 3.55 (m, 2H), 3.39 (d, *J* = 13.1 Hz, 1H), 3.26 (d, *J* = 13.1 Hz, 1H), 2.85 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.0, 143.2, 134.5, 132.4 (two peaks), 130.6 (two peaks), 130.1, 129.0, 128.2, 127.7, 126.8, 124.6, 122.6, 115.2, 114.9, 108.0, 56.3, 42.3, 33.3, 26.0. HRMS (ESI) *m/z* 378.1326 (M+H⁺), calcd for C₂₃H₂₀FNOS 378.1328. The *ee* was determined by HPLC analysis. LUX AMYLOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.7 min (major) and 6.3 min (minor).

4bc, **(5)**-(–)-**3**-**Benzyl**-**3**-(**(4**-**chlorobenzyl**)**thio**)-1-**methylindolin-2-one.** Colorless oil, 33 mg (0.1 mmol), 84% yield, 90% ee. $[\alpha]_{26}^{26}$ –72.0 (*c* 0.30, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.30 (d, *J* = 7.3 Hz, 1H), 7.25–7.13 (m, 3H), 7.12–6.98 (m, 6H), 6.87 (d, *J* = 6.4 Hz, 2H), 6.66–6.42 (m, 1H), 3.74–3.52 (m, 2H), 3.38 (d, *J* = 13.1 Hz, 1H), 3.25 (d, *J* = 13.1 Hz, 1H), 2.83 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 175.9, 143.2, 135.3, 134.5, 132.8, 130.4 (two peaks), 129.0, 128.4, 128.1, 127.7, 126.9, 124.7, 122.6, 108.1, 56.3, 42.3, 33.4, 26.0. HRMS (ESI) *m/z* 394.1035 (M+H⁺), calcd for C₂₃H₂₀ClNOS 394.1032. The *ee* was determined by HPLC analysis. LUX AMYLOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.8 min (major) and 19.7 min (minor).

4bd, **(5)**-(–)-**3**-**Benzyl**-**1**-**methyl**-**3**-((**4**-**methylbenzyl**)**thio**)indolin-2-one. Colorless oil, 32 mg (0.1 mmol), 86% yield, 95% ee. $[α]_{26}^{26}$ -36.9 (*c* 0.26, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.57 (d, *J* = 7.3 Hz, 1H), 7.47 (dd, *J* = 14.9, 7.4 Hz, 2H), 7.32 (dd, *J* = 13.5, 6.6 Hz, 4H), 7.26 (s, 3H), 7.14 (d, *J* = 6.6 Hz, 2H), 6.81 (d, *J* = 7.8 Hz, 1H), 3.87 (q, *J* = 12.4 Hz, 2H), 3.66 (d, *J* = 13.1 Hz, 1H), 3.52 (d, *J* = 13.1 Hz, 1H), 3.11 (s, 3H), 2.52 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 143.3, 136.7, 134.7, 133.5, 130.2, 128.9 (two peaks), 128.4, 127.7, 126.8, 124.6, 122.5, 108.0, 56.3, 42.2, 33.6, 26.0, 21.1. HRMS (ESI) *m*/*z* 374.1578 (M+H⁺), calcd for C₂₄H₂₃NOS 374.1579. The *ee* was determined by HPLC analysis. LUX AMYLOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.6 min (major) and 16.9 min (minor). **4be**, (**5**)-(–)-**3**-**Benzyl-3**-((**4**-(*tert*-**butyl**)**benzyl**)**thio**)-**1**-**methylindolin-2-one.** Colorless oil, 34 mg (0.1 mmol), 83% yield, 90% ee. $[\alpha]_{D}^{26}$ –27.6 (*c* 0.70, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.31 (d, *J* = 7.4 Hz, 1H), 7.22 (dd, *J* = 16.6, 8.7 Hz, 3H), 7.05 (t, *J* = 11.0 Hz, 6H), 6.89 (d, *J* = 6.7 Hz, 2H), 6.55 (d, *J* = 7.7 Hz, 1H), 3.71–3.51 (m, 2H), 3.42 (d, *J* = 13.1 Hz, 1H), 3.28 (d, *J* = 13.1 Hz, 1H), 2.83 (s, 3H), 1.26 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 176.0, 149.9, 143.3, 134.7, 133.5, 130.2, 128.8 (two peaks), 128.3, 127.7, 126.8, 125.2, 124.6, 122.5, 108.0, 56.3, 42.2, 34.5, 33.6, 31.3, 26.0 HRMS (ESI) *m/z* 416.2049 (M+H⁺), calcd for C₂₇H₂₉NOS 416.2048. The *ee* was determined by HPLC analysis. LUX AMYLOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; low rate 1.0 mL/min; 25 °C; 254 nm; retention time: 5.5 min (major) and 6.1 min (minor).

4bf, **(S)**-(–)-**3**-**Benzyl**-**1**-methyl-**3**-(**(3**-methylbenzyl)thio)indolin-2-one. Colorless oil, 29 mg (0.1 mmol), 78% yield, 92% ee. $[α]_{26}^{D}$ –27.8 (*c* 0.88, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.33 (d, *J* = 7.3 Hz, 1H), 7.22 (dd, *J* = 15.0, 7.3 Hz, 2H), 7.16–7.00 (m, SH), 6.97 (d, *J* = 7.5 Hz, 1H), 6.91 (d, *J* = 6.9 Hz, 3H), 6.56 (d, *J* = 7.7 Hz, 1H), 3.74–3.50 (m, 2H), 3.42 (d, *J* = 13.0 Hz, 1H), 3.27 (d, *J* = 13.0 Hz, 1H), 2.85 (*s*, 3H), 2.27 (*s*, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.1, 143.3, 137.9, 136.4, 134.7, 130.1, 129.8, 128.9, 128.2 (two peaks), 127.8 (two peaks), 126.8, 126.1, 124.6, 122.5, 108.0, 56.3, 42.2, 34.0, 26.0, 21.3. HRMS (ESI) *m*/*z* 374.1581 (M+H⁺), calcd for C₂₄H₂₃NOS 374.1579. The *ee* was determined by HPLC analysis. CHIRALPAK AD-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 14.0 min (major) and 12.3 min (minor).

4bg, **(S)-(–)-3-Benzyl-1-methyl-3-((2-methylbenzyl)thio)**indolin-2-one. Colorless oil, 28 mg (0.1 mmol), 76% yield, 92% ee. $[α]_{26}^{D}$ –33.2 (*c* 0.34, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.35 (d, *J* = 7.3 Hz, 1H), 7.22 (t, *J* = 7.7 Hz, 1H), 7.16–7.00 (m, 8H), 6.91 (d, *J* = 5.1 Hz, 2H), 6.60 (d, *J* = 7.8 Hz, 1H), 3.75 (d, *J* = 11.4 Hz, 1H), 3.60 (d, *J* = 11.4 Hz, 1H), 3.47 (d, *J* = 13.1 Hz, 1H), 3.29 (d, *J* = 13.1 Hz, 1H), 2.96 (s, 3H), 2.27 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.0, 143.2, 137.1, 134.8, 134.1, 130.3 (two peaks), 128.9, 128.5, 127.6 (two peaks), 126.8, 125.9, 124.6, 122.6, 108.0, 56.4, 42.0, 31.6, 26.1, 19.0 HRMS (ESI) *m/z* 374.1570 (M+H⁺), calcd for C₂₄H₂₃NOS 374.1579. The ee was determined by HPLC analysis. CHIRALCEL OD-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.2 min (major) and 5.4 min (minor).

4bh, **(S**)-(–)-3-Benzyl-3-(cyclohexylthio)-1-methylindolin-2one. Colorless oil, 23 mg (0.1 mmol), 65% yield, 86% ee. $[α]_D^{26}$ -40.1 (*c* 0.34, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.35 (d, *J* = 7.4 Hz, 1H), 7.19 (dt, *J* = 7.7, 3.8 Hz, 1H), 7.14–6.99 (m, 4H), 6.86 (dd, *J* = 7.3, 1.9 Hz, 2H), 6.58 (d, *J* = 7.7 Hz, 1H), 3.41 (d, *J* = 12.9 Hz, 1H), 3.24 (d, *J* = 13.0 Hz, 1H), 2.98 (s, 3H), 2.64 (s, 1H), 1.79 (d, *J* = 13.0 Hz, 1H), 1.72–1.52 (m, 4H), 1.48–1.31 (m, 2H), 1.17 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.9, 142.9, 134.7, 130.1, 129.4, 128.7, 127.6, 126.7, 124.5, 122.5, 107.9, 56.2, 43.2, 42.3, 35.1, 34.1, 26.2, 25.8 (two peaks), 25.4. HRMS (ESI) *m*/*z* 352.1732 (M+H⁺), calcd for C₂₂H₂₅NOS 352.1735.The ee was determined by HPLC analysis. CHIRALCEL OZ-H (4.6 mm i.d. × 250 mm); hexane/2propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.0 min (major) and 23.3 min (minor).

40a, (5)-(–)-3-(Benzylthio)-1,3-dimethylindolin-2-one. Colorless oil, 24 mg (0.1 mmol), 85% yield; 90% ee. $[\alpha]_D^{26}$ –56.6 (*c* 0.55, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.27 (m, 2H), 7.24–7.05 (m, 6H), 6.79 (d, *J* = 7.8 Hz, 1H), 3.59 (s, 2H), 3.06 (s, 3H), 1.66 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 136.64, 130.9, 128.9 (two peaks), 128.3, 127.1, 123.7, 123.0 108.2, 50.7, 34.2, 26.3, 22.6. HRMS (ESI) *m/z* 284.1107 (M+H⁺), calcd for C₁₇H₁₇NOS 284.1109. The ee was determined by HPLC analysis. LUX CELLULOSE-2 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 0.5 mL/min; 25 °C; 254 nm; retention time: 11.7 min (major) and 12.6 min (minor).

4pa, (5)-(–)-3-(Benzylthio)-3-ethyl-1-methylindolin-2-one. Colorless oil, 21 mg (0.1 mmol), 70% yield, 92% ee. $[\alpha]_D^{26}$ –114.7 (*c* 0.16, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.31 (t, *J* = 7.9 Hz, 2H), 7.22–7.06 (m, 6H), 6.79 (d, *J* = 7.7 Hz, 1H), 3.59 (s, 2H), 3.07 (s, 3H), 2.24–1.98 (m, 2H), 0.69 (t, J = 7.4 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 143.6, 136.8, 128.9 (two peaks), 128.3, 127.0, 124.1, 122.9 9, 108.1, 55.6, 33.7, 29.4, 26.2, 23.5, 9.0. HRMS (ESI) m/z 297.1186 (M+H⁺), calcd for C₁₈H₁₉NOS 297.1187. The ee was determined by HPLC analysis. CHIRALPAK IB (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.5 min (major) and 13.3 min (minor).

4ob, **(S)**-(–)-**3**-((**4**-Fluorobenzyl))hio)-1,3-dimethylindolin-2one. Colorless oil, 24 mg (0.1 mmol), 81% yield, 92% ee. $[\alpha]_D^{26}$ -78.0 (*c* 0.39, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.42–7.22 (m, 2H), 7.20–7.00 (m, 3H), 6.88 (t, *J* = 8.2 Hz, 2H), 6.80 (d, *J* = 7.9 Hz, 1H), 3.57 (s, 2H), 3.08 (s, 3H), 1.65 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 177.3, 163.5, 160.2, 142.7, 132.4, 130.9–130.3, 129.0, 123.7, 123.0, 115.2, 115.0, 108.3, 50.8, 33.4, 26.3, 22.7. HRMS (ESI) *m/z* 302.1016 (M+H⁺), calcd for C₁₇H₁₆FNOS 302.1015. The ee was determined by HPLC analysis. CHIRALCEL OZ-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 15.0 min (major) and 17.1 (minor).

40e, **(5)**-(–)-**3**-((**4**-(*tert*-**Buty**])**b**enzy])**thio**)-**1**,**3**-dimethylindolin-2-one. Colorless oil, 31 mg (0.1 mmol), 93% yield, 92% ee. $[\alpha]_{2^6}^{D}$ -36.5 (*c* 1.20, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.41–7.18 (m, 4H), 7.10 (t, *J* = 7.3 Hz, 1H), 7.02 (d, *J* = 7.6 Hz, 2H), 6.78 (d, *J* = 7.7 Hz, 1H), 3.57 (s, 2H), 3.05 (s, 3H), 1.67 (s, 3H), 1.27 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 177.4, 149.9, 142.8, 133.5, 131.0, 128.7 (two peaks), 125.2, 123.7, 122.9, 108.2, 50.7, 34.5, 33.8, 31.3, 26.3, 22.7. HRMS (ESI) *m/z* 340.1732 (M+H⁺), calcd for C₂₁H₂₅NOS 340.1735. The ee was determined by HPLC analysis. CHIRALCEL OZ-H 4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 11.7 (major) and 13.2 (minor).

4oh, **(S)-(–)-3-(Cyclohexylthio)-1,3-dimethylindolin-2-one.** Colorless oil, 18 mg (0.1 mmol), 65% yield, 92% ee. $[\alpha]_D^{26}$ –94.0 (*c* 0.10, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.40–7.29 (m, 2H), 7.11 (dd, *J* = 11.2, 3.8 Hz, 1H), 6.91–6.78 (m, 1H), 3.24 (s, 3H), 2.57 (s, 1H), 1.86–1.73 (m, 1H), 1.65 (s, 3H), 1.57 (d, *J* = 7.9 Hz, 3H), 1.51–1.28 (m, 3H), 1.18 (dd, *J* = 13.3, 7.0 Hz, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 178.3, 142.4, 132.1, 128.7, 123.7, 123.0, 108.2, 50.5, 42.4, 35.2, 34.2, 26.5, 25.9 (two peaks), 25.5, 23.4. HRMS (ESI) *m/z* 276.1428 (M+H⁺), calcd for C₁₆H₂₁NOS 276.1422. The ee was determined by HPLC analysis. CHIRALCEL OZ-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 16.4 (major) and 19.1 (minor).

5aa, (S)-(+)-3-Benzyl-3-(benzylthio)benzofuran-2(3*H***)-one. White solid, mp 98.5–100.4 °C, 30 mg (0.1 mmol), 87% yield, 90% ee. [\alpha]_{D}^{26} +21.31 (***c* **0.40, CHCl₃). ¹H NMR (300 MHz, CDCl₃) \delta 7.35–7.17 (m, 8H), 7.17–7.09 (m, 3H), 7.01–6.89 (m, 3H), 3.84 (d,** *J* **= 12.2 Hz, 1H), 3.70 (d,** *J* **= 12.2 Hz, 1H), 3.52 (d,** *J* **= 13.4 Hz, 1H), 3.38 (d,** *J* **= 13.4 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) \delta 175.8, 152.6, 135.8, 133.9, 130.1, 129.8, 129.2, 128.6, 128.2, 127.4 (two peaks), 126.9, 124.8, 124.4, 110.9, 55.2, 42.3, 34.5. HRMS (EI)** *m/z* **346.1030 (M⁺), calcd for C₂₂H₁₈O₂S 346.1028, The** *ee* **was determined by HPLC analysis. CHIRALPAK IE (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 10.1 min (minor) and 10.8 min (major).**

5ba, **(S)-3-(Benzylthio)-3-(4-fluorobenzyl)benzofuran-2(3***H***)one. White solid, mp 123.5–124.8 °C, 25 mg (0.1 mmol), 68% yield, 71% ee. [\alpha]_D^{26} +10.34 (***c* **0.60, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.26–7.20 (m, 2H), 7.19–7.06 (m, 6H), 6.87–6.77 (m, 3H), 6.72 (t,** *J* **= 8.7 Hz, 2H), 3.72 (d,** *J* **= 12.2 Hz, 1H), 3.59 (d,** *J* **= 12.2 Hz, 1H), 3.38 (d,** *J* **= 13.5 Hz, 1H), 3.24 (d,** *J* **= 13.5 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 175.7, 163.7, 160.4, 152.6, 135.7, 131.7 (two peaks), 130.0, 129.6, 129.2, 128.6, 127.5, 126.7, 124.6 (two peaks), 115.3, 115.0, 111.0, 55.2, 41.5, 34.5. HRMS (EI)** *m/z* **364.0930 (M⁺), calcd for C₂₂H₁₇FO₂S 364.0933. The** *ee* **was determined by HPLC analysis. CHIRALPAK IC–IC (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 19.9 min (major) and 27.0 min (minor).**

5ca, (S)-(+)-3-(Benzylthio)-3-(3-fluorobenzyl)benzofuran-2(3H)-one. White solid, mp 87.7–89.2 °C, 31 mg (0.1 mmol), 86% yield, 95% ee. $[\alpha]_{26}^{26}$ +22.00 (c 0.30, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.26–7.07 (m, 8H), 7.00 (td, *J* = 7.9, 6.2 Hz, 1H), 6.85 (d, *J* = 8.0 Hz, 1H), 6.75 (td, *J* = 8.4, 2.2 Hz, 1H), 6.65 (d, *J* = 7.7 Hz, 1H), 6.55 (dd, *J* = 9.7, 1.8 Hz, 1H), 3.73 (d, *J* = 12.2 Hz, 1H), 3.61 (d, *J* = 12.2 Hz, 1H), 3.40 (d, *J* = 13.4 Hz, 1H), 3.26 (d, *J* = 13.4 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 175.5, 164.0, 160.7, 152.6, 136.3 (two peaks), 135.7, 130.1, 129.7 (two peaks), 129.2, 128.6, 127.6, 126.6, 125.9 (two peaks), 124.6 (two peaks), 117.2, 116.9, 114.6, 114.3, 111.0, 54.9, 41.9 (two peaks), 34.5. HRMS (EI) *m*/*z* 364.0932 (M⁺), calcd for C₂₂H₁₇FO₂S 364.0933. The *ee* was determined by HPLC analysis. CHIRALPAK IC–IC (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 18.9 min (major) and 21.2 min (minor).

5da, **(S)**-(+)-**3**-(**Benzylthio**)-**3**-(**2**-fluorobenzyl)benzofuran-**2**(*3H*)-one. White solid, mp 95.1–96.8 °C, 34 mg (0.1 mmol), 93% yield, 91% ee. $[\alpha]_{26}^{26}$ +26.00 (*c* 0.40, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.27–7.19 (m, 1H), 7.20–7.08 (m, 6H), 7.08–6.97 (m, 3H), 6.90–6.80 (m, 2H), 6.74 (t, *J* = 9.1 Hz, 1H), 3.76 (d, *J* = 12.1 Hz, 1H), 3.63 (d, *J* = 12.1 Hz, 1H), 3.51 (d, *J* = 13.7 Hz, 1H), 3.39 (d, *J* = 13.7 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 175.8, 162.4, 159.2, 152.5, 135.7, 131.6 (two peaks), 129.9, 129.5, 129.4 (two peaks), 129.2, 128.6 (two peaks), 127.5, 126.5, 125.2 (two peaks), 124.4, 124.1 (two peaks), 121.6 (two peaks), 115.5, 115.2, 110.6, 54.6, 43.2, 34.5, 34.4 (two peaks). HRMS (EI) *m*/*z* 364.0932 (M⁺), calcd for C₂₂H₁₇FO₂S 364.0933. The ee was determined by HPLC analysis. CHIRACEL OD-H (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 7.6 min (minor) and 8.4 min (major).

5ea, (S)-(+)-3-(Benzylthio)-3-(4-methoxybenzyl)benzofuran-2(3*H***)-one.** yellow oil, 29 mg (0.1 mmol), 78% yield, 85% ee. $[\alpha]_D^{26}$ +44.88 (*c* 0.40, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.30 (t, *J* = 5.5 Hz, 1H), 7.20 (ddd, *J* = 14.2, 6.4, 4.6 Hz, 8H), 6.91 (d, *J* = 7.9 Hz, 1H), 6.84 (d, *J* = 8.6 Hz, 2H), 6.63 (d, *J* = 8.6 Hz, 2H), 3.78 (d, *J* = 12.2 Hz, 1H), 3.70 (s, 3H), 3.65 (d, *J* = 12.2 Hz, 1H), 3.42 (d, *J* = 13.5 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 175.9, 158.8, 152.7, 135.8, 131.2, 129.8, 129.2, 128.5, 127.4, 127.1, 125.9, 124.8, 124.4, 113.6, 110.9, 55.2 (two peaks), 41.5, 34.5, 29.7. HRMS (EI) *m/z* 376.1136 (M⁺), calcd for C₂₃H₂₀O₃S 376.1133. The ee was determined by HPLC analysis. CHIRALPAK IC (4.6 mm i.d. × 250 mm); hexane/2-propanol = 95/5; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 9.8 min (minor) and 11.6 min (major).

5fa, (5)-3-(Benzylthio)-3-methylbenzofuran-2(3*H***)-one.** Colorless oil, 21 mg (0.1 mmol), 78% yield, 88% ee. $[\alpha]_D^{26}$ +15.06 (*c* 0.64, CHCl₃), ¹H NMR (300 MHz, CDCl₃) δ 7.37–7.30 (m, 2H), 7.25–7.07 (m, 7H), 3.73 (d, *J* = 12.2 Hz, 1H), 3.63 (d, *J* = 12.2 Hz, 1H), 1.79 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.8, 152.3, 135.8, 129.8, 129.2 (two peaks), 128.5, 127.4, 124.8, 124.0, 111.0, 49.3, 34.6, 22.9. HRMS (ESI) *m/z* 271.0801 (M+H⁺), calcd for C₁₆H₁₅O₂S 271.0793. The ee was determined by HPLC analysis. CHIRALPAK IC–IC (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 21.7 min (minor) and 22.9 min (major).

5ai, (5)-(+)-3-Benzyl-3-(propylthio)benzofuran-2(3*H***)-one. White solid, mp 48.4–50.1 °C, 21 mg (0.1 mmol), 71% yield, 76% ee. [\alpha]_D^{26} +54.18 (***c* **0.50, CHCl₃). ¹H NMR (300 MHz, CDCl₃) \delta 7.24 (dt,** *J* **= 10.0, 4.9 Hz, 1H), 7.19–6.99 (m, 5H), 6.92–6.78 (m, 3H), 3.42 (d,** *J* **= 13.4 Hz, 1H), 3.27 (d,** *J* **= 13.4 Hz, 1H), 2.45 (dt,** *J* **= 11.7, 7.2 Hz, 1H), 2.30 (dt,** *J* **= 11.7, 7.5 Hz, 1H), 1.48–1.33 (m, 2H), 0.84 (t,** *J* **= 7.3 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) \delta 176.4, 152.5, 134.1, 130.1, 129.6, 128.2, 127.4 (two peaks), 124.8, 124.4, 110.7, 54.6, 42.3, 31.6, 22.1, 13.5. HRMS (EI)** *m/z* **298.1030 (M⁺), calcd for C₁₈H₁₈O₂S 298.1028. The ee was determined by HPLC analysis. LUX CELLULOSE-4 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 0.8 mL/min; 25 °C; 254 nm; retention time: 11.0 min (minor) and 12.3 min (major).**

5fi, (*S*)-(+)-3-Methyl-3-(propylthio)benzofuran-2(3*H*)-one. Colorless oil, 20 mg (0.1 mmol), 91% yield, 74% ee. $[\alpha]_D^{26}$ +35.01 (*c* 0.40, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.26 (s, 1H), 7.25–7.01 (m, 3H), 2.49–2.19 (m, 2H), 1.71 (s, 3H), 1.40 (dd, *J* = 14.1, 7.0 Hz, 2H), 0.83 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 176.3, 151.2, 128.9, 128.6, 123.7, 122.8, 109.8, 47.6, 30.7, 21.8, 21.0,

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12.4. HRMS (EI) m/z 222.0719 (M⁺), calcd for C₁₂H₁₄O₂S 222.0715. The ee was determined by HPLC analysis. LUX CELLULOSE-4 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 99/1; flow rate 0.8 mL/min; 25 °C; 254 nm; retention time: 7.4 min (minor) and 8.0 min (major).

6, (**5**)-(+)-**3**-Benzyl-**3**-((**4**-(*tert*-butyl)benzyl)sulfonyl)-**1**-methylindolin-**2**-one. Colorless oil, 95% yield, 90% ee. $[\alpha]_D^{26}$ +33.8 (*c* 0.31, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.63 (d, *J* = 7.5 Hz, 1H), 7.32–7.21 (m, 4H), 7.19 (d, *J* = 6.2 Hz, 1H), 7.04 (t, *J* = 7.6 Hz, 1H), 7.00–6.89 (m, 3H), 6.80 (dd, *J* = 7.3, 1.6 Hz, 2H), 6.52 (d, *J* = 7.8 Hz, 1H), 4.54 (d, *J* = 13.4 Hz, 1H), 4.42 (d, *J* = 13.4 Hz, 1H), 3.73 (d, *J* = 12.8 Hz, 1H), 3.55 (d, *J* = 12.8 Hz, 1H), 2.90 (s, 3H), 1.22 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 170.4, 152.0, 144.4, 132.7, 131.2, 130.6, 129.9, 128.0, 127.2, 126.7, 125.7, 123.1 (two peaks), 121.4, 108.5, 75.2, 54.9, 37.3, 34.7, 31.2. HRMS (ESI) *m/z* 447.1869 (M +H⁺), calcd for C₂₇H₂₉O₃NS 447.1868. The ee was determined by HPLC analysis. LUX CELLULOSE-1 (4.6 mm i.d. × 250 mm); hexane/2-propanol = 90/10; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 20.1 (major) and 26.8 (minor).

7, **(5)**-(+)-3-Benzyl-3-(*p*-tolylthio)benzofuran-2(3*H*)-one. White solid, mp 125.4–128.7, 25 mg (0.1 mmol), 72% yield; 75% ee. $[\alpha]_D^{26}$ +12.06 (*c* 0.70, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.43–7.36 (m, 1H), 7.22–7.04 (m, 7H), 6.97 (dd, *J* = 10.3, 6.1 Hz, 4H), 6.72–6.63 (m, 1H), 3.51 (d, *J* = 13.4 Hz, 1H), 3.43 (d, *J* = 13.4 Hz, 1H), 2.28 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 175.5, 152.6, 140.4, 136.6, 134.2, 130.1, 129.6 (two peaks), 128.2, 127.3, 125.1 (two peaks), 124.1, 110.6, 58.9, 41.7, 21.3; HRMS (ESI) *m/z* 347.1109 (M +H⁺), calcd for C₂₂H₁₉O₂S 347.1106. The *ee* was determined by HPLC analysis. CHIRALPAK IC-IC (4.6 mm i.d. × 250 mm); Hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 25.9 min (major) and 28.3 min (minor).

8, (*S*)-(+)-3-Methyl-3-(*p*-tolylthio)benzofuran-2(3*H*)-one. White solid, mp 73.5–76.0, 20 mg (0.1 mmol), 72% yield, 75% ee. $[\alpha]_D^{26}$ +11.94 (*c* 0.60, CHCl₃). ¹H NMR (300 MHz, CDCl₃) δ 7.38 (dd, *J* = 7.3, 1.5 Hz, 1H), 7.25–7.07 (m, 4H), 6.98 (d, *J* = 7.9 Hz, 2H), 6.91–6.82 (m, 1H), 2.28 (s, 3H), 1.78 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 176.6, 152.2, 140.4, 129.6 (three peaks), 125.7, 124.4, 124.1, 110.6, 53.1, 22.0, 21.3; HRMS (ESI) *m*/*z* 271.0791 (M+H⁺), calcd for C₁₆H₁₅O₂S 271.0793. The *ee* was determined by HPLC analysis. LUX CELLULOSE-4 (4.6 mm i.d. × 250 mm); Hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 10.6 min (minor) and 11.9 min (major).

9, **(5)-3-(Benzylthio)-3-phenylbenzofuran-2(3***H***)-one.** Colorless oil, 23 mg (0.1 mmol), 69% yield, 0% ee. ¹H NMR (300 MHz, CDCl₃) δ 7.61 (d, J = 7.2 Hz, 2H), 7.30 (dt, J = 11.8, 7.2 Hz, 5H), 7.11 (ddd, J = 8.4, 7.7, 5.0 Hz, 7H), 3.71 (d, J = 11.8 Hz, 1H), 3.55 (d, J = 11.8 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 174.8, 152.2, 135.4 (two peaks), 130.0, 129.2 (two peaks), 128.5 (three peaks), 127.5, 125.5, 124.8, 111.2, 57.2, 35.4; HRMS (ESI) *m/z* 333.0946 (M+H⁺), calcd for C₂₁H₁₇O₂S 333.0949. The *ee* was determined by HPLC analysis. LUX CELLULOSE-2 (4.6 mm i.d. × 250 mm); Hexane/2-propanol = 99/1; flow rate 1.0 mL/min; 25 °C; 254 nm; retention time: 8.9 and 10.6 min.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b01606.

Copies of HPLC spectra, compounds 7–9, determination of the absolute configuration by X-ray crystallography and copies of NMR spectra (PDF) More experimental data (ZIP)

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Notes

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(16) CCDC1407355 (**5aa**) contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam. ac.uk/data_request/cif.